

# A New Rhodamine-Based “Off-On” Fluorescent Chemosensor for Hg (II) Ion and its Application in Imaging Hg (II) in Living Cells

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**Abstract** A novel rhodamine derivative (Rh-C), synthesized by the reaction of rhodamine ethylenediamine and cinnamoyl chloride, was evaluated as a chemoselective  $\text{Hg}^{2+}$  ion sensor. Addition of  $\text{Hg}^{2+}$  to an ethanol aqueous solution of the Rh-C resulted in a color change from colorless to obvious pink color together with distinctive changes in UV–vis absorption spectrum and fluorescence spectrum. However, other common alkali-, alkaline earth-, transition- and rare earth metal ions induced no or minimal spectral changes. The interaction of  $\text{Hg}^{2+}$  and sensor Rh-C was proven to adopt a 1:1 binding stoichiometry and the recognition process is reversible. The chemosensor displayed a linear response to  $\text{Hg}^{2+}$  in the range of 0.4–5  $\mu\text{M}$  with a detection limit of  $7.4 \times 10^{-8}$  M. The sensor Rh-C was also successfully applied to the imaging of  $\text{Hg}^{2+}$  in HL-7702 cells.

**Keywords** Hg (II) ion · Rhodamine derivative · Fluorescence enhancement · Chemosensor · Fluorescent image · living cells

## Introduction

Mercury is a highly toxic and widespread global pollutant [1–4]. The mercuric ion, Hg [II], combines with both inorganic and organic ligands, which can readily penetrate through biological membranes even at very low concentrations [5, 6]. A wide variety of symptoms, including central

nervous system, kidney, and endocrine system diseases result from a series of biological effects [7–9]. Thus, much attention has been focused on developing  $\text{Hg}^{2+}$  fluorescent chemosensors with excellent sensitivity and selectivity, quick response time and easy signal detection [10–16].

The rhodamine-based chemosensors can react specifically with specific metal ions to induce a concomitant change of their photochemical properties (excitation/emission wavelength, fluorescence intensity, and so forth). Alteration in molecular structure between non-fluorescent of spirocyclic and fluorescent ring-open conformations of rhodamine framework is employed as the detection mechanism [17–22].

Since  $\text{Hg}^{2+}$  is a heavy metal ion with  $5d^{10} 6s^0$  electronic configuration, the oxygen atom or the nitrogen atom of the imino moiety might be a proper binding site when incorporated into rhodamine fluorophore [23–27]. The increase of the O, N atom has the potential to enhance the selective and sensitivity of the resulting chemosensors toward  $\text{Hg}^{2+}$ . Herein, we synthesized a rhodamine-based fluorogenic chemosensor (Rh-C), for selective response to  $\text{Hg}^{2+}$  in aqueous media and in living cells. Further, upon chelation of  $\text{Hg}^{2+}$ , Rh-C will change to a fluorescent ring-opened form, which could be detected by the naked eye.

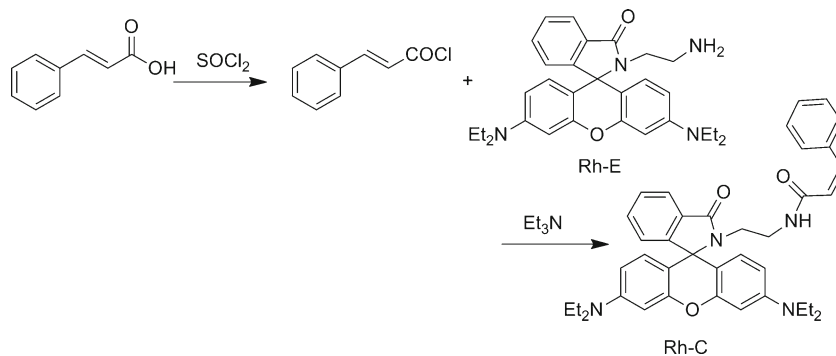
## Experiments

### Materials and Instruments

All the reagents were purchased from commercial suppliers and used without further purification. All the common chemicals were of analytical grade. Solvents were purified by standard procedures. All reactions were monitored by TLC (thin-layer chromatography) with detection by UV. The solutions of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Ba}^{2+}$ ,  $\text{Fe}^{3+}$ ,

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**Scheme 1** Synthetic routes of the probe Rh-C

$\text{Zn}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Ag}^+$  and  $\text{Hg}^{2+}$  were prepared from their nitrate salts. Doubly distilled deionized water was used throughout the experiment.

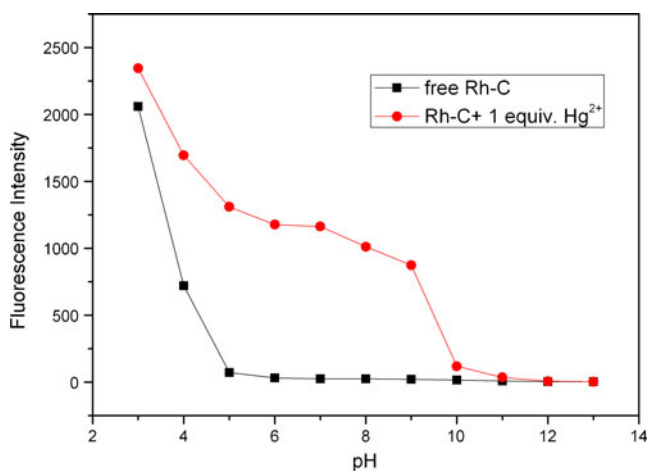
The absorption spectra were acquired on a Purkinje General TU-1901 UV–vis spectrometer. Fluorescence spectra measurements were performed on a Hitachi F-4500 Fluorescence spectrophotometer at room temperature. NMR spectra were measured on a Bruker 300 MHz spectrometer with chemical shifts reported in ppm (in  $\text{CDCl}_3$ ; TMS as internal standard). Electrospray ionization (ESI) mass spectra were conducted by ABI-057-TY4675 instrument. The pH measurements were carried out on a PHS-3W pH meter. The masses of the samples and solvents were determined by a Mettler Toledo AB204-N analytical balance with an accuracy of 0.0001 g. Fluorescence images experiments were carried out with an Olympus IX71 inverted fluorescence microscope.

#### Synthesis of Probe Rh-C

The newly synthesized rhodamine B derivative (Rh-C) was prepared in high yield (Scheme 1). Compound rhodamine

ethylenediamine (Rh-E) was facilely synthesized in high yield according to the procedure as published in the literature [28].

Rhodamine ethylenediamine (Rh-E) (2.52 g, 5.19 mmol) and triethylamine (4 mL) were dissolved in 25 mL dry dichloromethane. After cooling to 0 °C in ice bath, to the solution was added dropwise a solution of acinnamoyl chloride (0.86 g, 5.19 mmol) in 20 mL of dichloromethane over 10 min. The resulting mixture was stirred for 2 h at room temperature. The reaction mixture was then evaporated and the crude product was purified by column chromatography (silica gel, petroleum ether: ethyl acetate 1: 1, v/v). The yield was 72 %.  $^1\text{H}$ NMR( $\text{CDCl}_3$ , 300 MHz):  $\delta$ =7.98 (dd, 1H), 7.57 (s, 1H), 7.51 (m, 2H), 7.47 (dd, 2H), 7.35 (m, 3H), 7.08 (dd, 2H), 6.47 (d,  $J$ =8.9Hz, 2H), 6.30–6.39(m, 5H), 3.18–3.34 (m, 12H), 1.19 (t,  $J$ =7.1Hz, 12H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz):  $\delta$ =170.15, 165.82, 153.85, 153.28, 140.10, 135.18, 132.84, 130.40, 129.32, 128.66, 128.45, 128.21, 127.84, 123.93, 122.83, 121.38, 108.39, 97.89, 65.82, 44.43, 41.24, 40.17, 12.56. IR (KBr,  $\nu/\text{cm}^{-1}$ ): 3427, 3006, 2854, 1672, 1615, 1459, 1220, 1118, 979. ESI-MS ( $M+\text{H}^+$ ):  $m/z$ =614.33 ( $\text{C}_{39}\text{H}_{42}\text{N}_4\text{O}_3$ ).

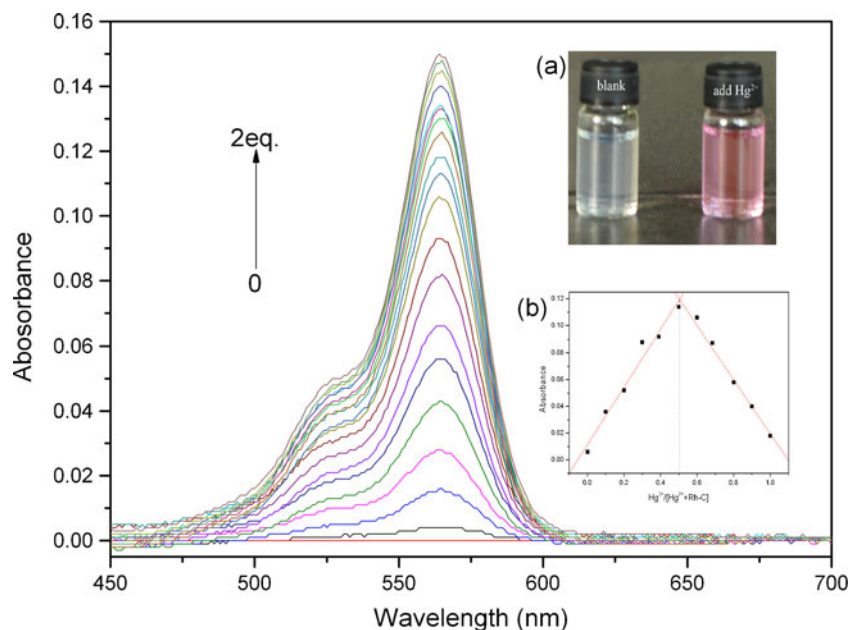


**Fig. 1** Fluorescence intensity of Rh-C (10  $\mu\text{M}$ ) in the absence and presence of 1 equiv.  $\text{Hg}^{2+}$  in water-ethanol solution (30:70, v/v) at different pH. The pH modified by adding 75 %  $\text{HClO}_4$  or  $\text{NaOH}$  (10 %). Excitation: 535 nm, emission: 586 nm

#### General UV–vis and Fluorescence Spectra Measurements

2 mM of each inorganic salt ( $\text{NaNO}_3$ ,  $\text{KNO}_3$ ,  $\text{Mg}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ ,  $\text{Mn}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ ,  $\text{Ca}(\text{NO}_3)_2$ ,  $\text{Ba}(\text{NO}_3)_2$ ,  $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ ,  $\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ ,  $\text{Pb}(\text{NO}_3)_2$ ,  $\text{Cu}(\text{NO}_3)_2 \cdot 3\text{H}_2\text{O}$ ,  $\text{Cd}(\text{NO}_3)_2 \cdot 2\text{H}_2\text{O}$ ,  $\text{AgNO}_3$  and  $\text{Hg}(\text{NO}_3)_2 \cdot 0.5\text{H}_2\text{O}$ ) was dissolved in distilled water to afford  $2 \times 10^{-3} \text{ mol} \cdot \text{L}^{-1}$  aqueous solution. A  $2.0 \times 10^{-3} \text{ mol} \cdot \text{L}^{-1}$  stock solution of Rh-C was prepared in absolute ethanol. All the measurements were made according to the following procedure. To 10 mL glass tubes containing different amounts of metal ions, proper amounts of the solution of Rh-C was added directly with micropipette, then diluted with buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v) to 10 mL, then the absorption and fluorescence sensing of metal ions were run. In selectivity experiments, the test samples were prepared by placing the appropriate amounts of metal ion stock solution into 10 mL solution of Rh-C (10  $\mu\text{M}$ ). All samples were prepared at room temperature, shaken for 10 s and waited for 10 min before test. Fluorescence measurements

**Fig. 2** UV–vis absorption spectra of Rh-C (10.0  $\mu\text{M}$ ) obtained during the titration by  $\text{Hg}^{2+}$  (0–2 equiv.) in buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v) at room temperature. Inset: **a** The color change of Rh-C (10  $\mu\text{M}$ )+2 equiv.  $\text{Hg}^{2+}$  in solution, **b** Job's plot of changes of absorbance at 562 nm, the total concentration of  $[\text{Hg}^{2+}]+[\text{Rh-C}]$  was 20  $\mu\text{M}$



were carried out with excitation and emission slit width of 10 and 5 nm and excitation wavelength was 535 nm.

#### Experimental Details for Cell Imaging Experiments

The HL-7702 cells (human hepatocyte cell line) were cultured in DEME (Invitrogen) supplemented with 10 % FBS (Invitrogen). One day before imaging, the cells were seeded in 6-well flat-bottomed plates. The next day, the HL-7702 cells were incubated with 10  $\mu\text{M}$  sensor Rh-C for 0.5 h at 37  $^{\circ}\text{C}$  in humidified environment of 5 %  $\text{CO}_2$  and then washed with phosphate-buffered saline (PBS) three times before incubating with 10  $\mu\text{M}$   $\text{Hg}(\text{NO}_3)_2$  for another 0.5 h, cells were rinsed with PBS three times again, then the

fluorescence imaging of intracellular  $\text{Hg}^{2+}$  was observed under inverted fluorescence microscope (IX71, Olympus, Japan) with a 40 $\times$  objective lens (excited with green light). The HL-7702 cells only incubated with 10  $\mu\text{M}$  Rh-C for 0.5 h at 37  $^{\circ}\text{C}$  under 5 %  $\text{CO}_2$  was as a blank control.

#### Determination of Binding Constants

The binding constant was calculated from the absorption intensity titration curves according to Benesi-Hildebrand equation as follow.

$$\frac{1}{A - A_0} = \frac{1}{K_a(A_{\text{max}} - A_0)[c]} + \frac{1}{A_{\text{max}} - A_0}$$

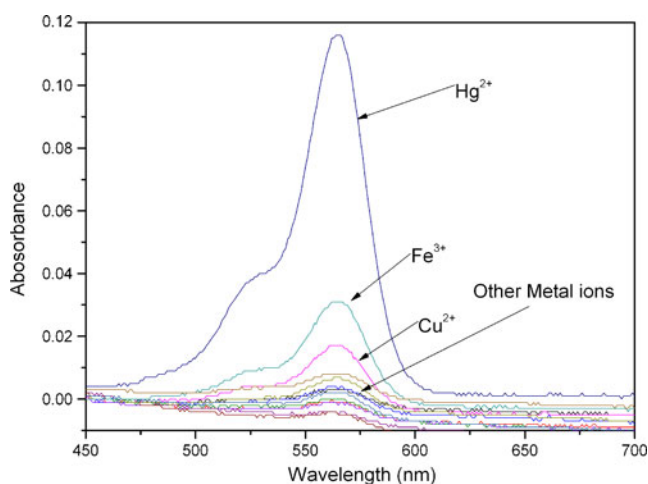
$A$  and  $A_0$  is the absorbance of Rh-C solution in the presence and absence of  $\text{Hg}^{2+}$ , respectively;  $A_{\text{max}}$  is the saturated absorbance of Rh-C in the presence of excess amount of  $\text{Hg}^{2+}$ ;  $[c]$  is the concentration of  $\text{Hg}^{2+}$  ions added ( $\text{mol L}^{-1}$ ). The association constant values  $K_a$  is calculate by the ratio intercept/slope.

## Results and discussion

### Synthesis and Structural Characterization of Rh-C

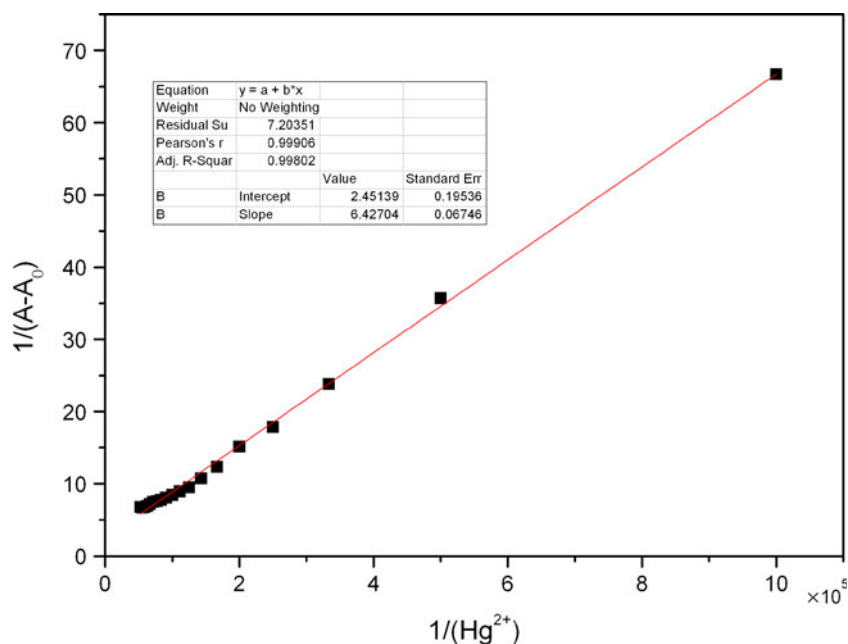
Rh-C was facilely synthesized from rhodamine B by acylation reaction, as summarized in Scheme 1. Its structure was confirmed by  $^1\text{H}$ NMR,  $^{13}\text{C}$ NMR and MS data.

A solution of Rh-C in ethanol is colorless and weakly fluorescent, indicating that the spiro lactam form of Rh-C



**Fig. 3** UV–vis spectra of Rh-C (10.0  $\mu\text{M}$ ) upon addition of 2.0 equiv. of  $\text{Hg}^{2+}$  and 10 equiv. other metal ions ( $\text{Ag}^+$ ,  $\text{Ba}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Pb}^{2+}$  and  $\text{Zn}^{2+}$ ) in buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v)

**Fig. 4** Benesi-Hildebrand plot (absorbance at 562 nm) of Rh-C using 1:1 stoichiometry for association between Rh-C and  $\text{Hg}^{2+}$



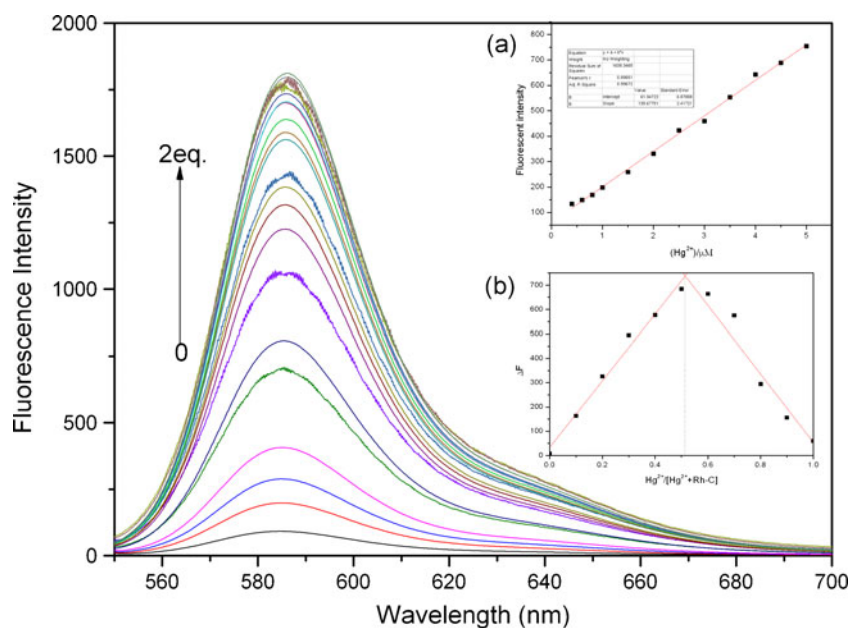
exists predominantly. The characteristic peak of the 9-carbon of Rh-C near 66 ppm in the  $^{13}\text{C}$ NMR spectrum also supports this consideration [29]. The chemosensing behavior of Rh-C was investigated by UV-vis and fluorescence measurements.

#### Effect of pH Value

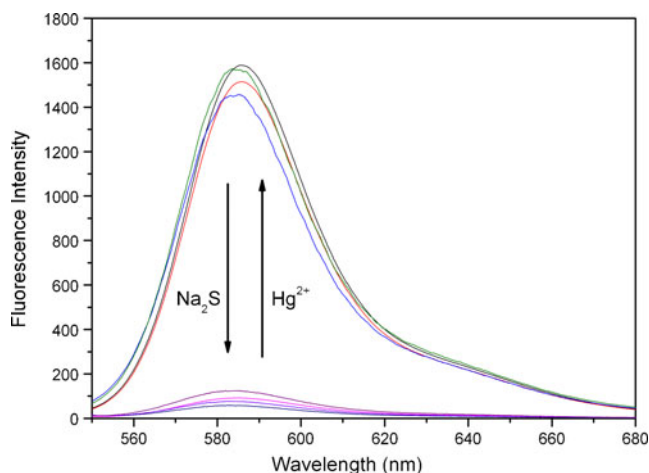
For practical application, the appropriate pH conditions for successful operation of the sensor were evaluated. The effects of pH on the fluorescence response of Rh-C obtained without and with  $\text{Hg}^{2+}$  in water-ethanol (7/3, v/v) are shown in Fig. 1. For free Rh-C, at acid conditions ( $\text{pH} < 5$ ), the ring

opening of rhodamine took place because of the strong protonation. No obvious fluorescence emission of Rh-C was observed between pH 5 and 12, suggesting that the compound is insensitive to pH and that the spirolactam form is still preferred in this condition. However, the addition of  $\text{Hg}^{2+}$  led to the fluorescence enhancement over a comparatively wide pH range (5.0–9.0), which is attributed to opening of the rhodamine ring. The results suggest that Rh-C was insensitive to pH near 7.0 and could work in physiological pH conditions with a very low background fluorescence [30]. Therefore, further UV-vis and fluorescent studies were carried out in buffered (HEPES 20 mM,  $\text{pH} = 7.0$ ) water-ethanol (7/3, v/v) at room temperature.

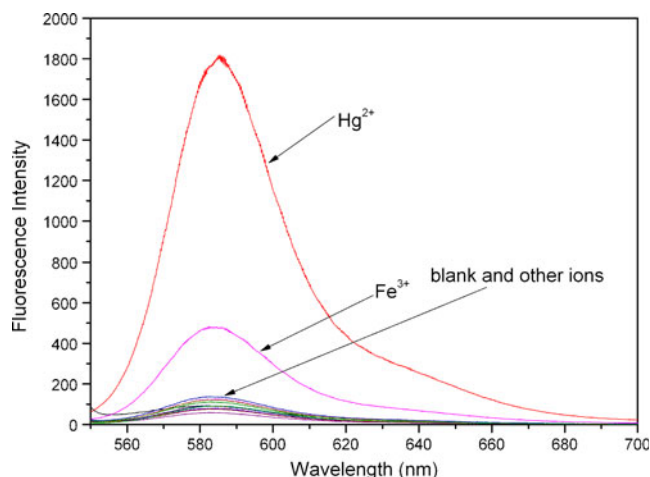
**Fig. 5** Fluorescence emission spectra of Rh-C (10.0  $\mu\text{M}$ ) in buffered (HEPES 20 mM,  $\text{pH} = 7.0$ ) water-ethanol (7/3, v/v) upon the addition of  $\text{Hg}^{2+}$  (0–2 equiv.) with an excitation of 535 nm. Inset: **a** fluorescence enhancement at 586 nm as a function of  $\text{Hg}^{2+}$  concentration, **b** Job's plot of changes of fluorescent intensity at 586 nm, the total concentration of  $[\text{Hg}^{2+}] + [\text{Rh-C}]$  was 10  $\mu\text{M}$







**Fig. 6** Reversibility of  $\text{Hg}^{2+}$  coordination to Rh-C (10  $\mu\text{M}$ ) by  $\text{Na}_2\text{S}$  in buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v). The top lines represent the fluorescence enhancement that occurs after addition of 1 equiv. of  $\text{Hg}^{2+}$ . The bottom lines represent the fluorescence intensity of free Rh-C and the fluorescence intensity decrease that occurs after addition 1 equiv. of  $\text{Na}_2\text{S}$  to a solution containing the  $[\text{Rh-C-Hg}^{2+}]$  species. Four cycles of on/off by  $\text{Hg}^{2+}/\text{Na}_2\text{S}$  addition are depicted in this plot.



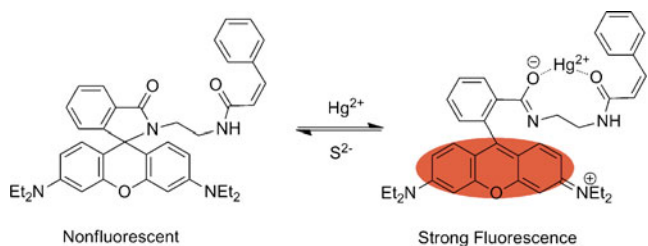
**Fig. 7** Fluorescent spectra of Rh-C (10.0  $\mu\text{M}$ ) upon addition of 2.0 equiv. of  $\text{Hg}^{2+}$  and 10 equiv. other metal ions ( $\text{Ag}^+$ ,  $\text{Ba}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Pb}^{2+}$  and  $\text{Zn}^{2+}$ ) in buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v)

UV–vis Spectral Responses of Rh-C

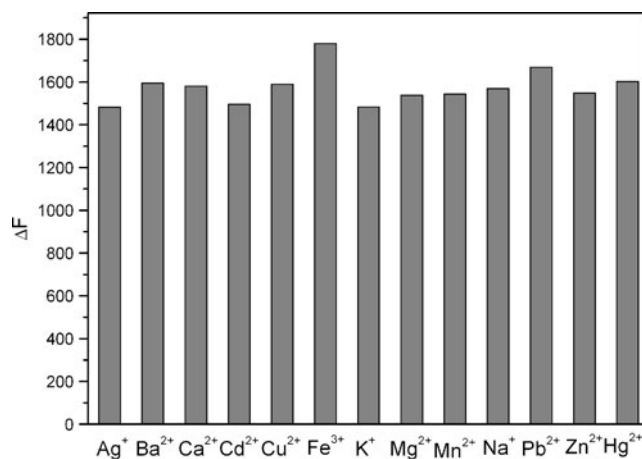
The absorption spectra of Rh-C with varying  $\text{Hg}^{2+}$  concentrations in buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v) were recorded, as shown in Fig. 2. Like most of the spirocycle RhB derivatives, When no  $\text{Hg}^{2+}$  was added to the solution, free Rh-C was colorless and exhibited almost no absorption peak in the visible wavelength range (>400 nm) due to the closed spirolactam ring. In the presence of 2 equiv. of  $\text{Hg}^{2+}$ , the absorbance was enhanced obviously and a new peak at 562 nm was observed, accompanied by a clear color change from colorless to pink. This enhancement in absorbance can be ascribed to the clear formation of the ring-opened amide form of Rh-C upon  $\text{Hg}^{2+}$  ions binding [19, 31]. The color of Rh-C also change from colorless to pink (Fig. 2.Inset (a)), indicating that Rh-C can serve as a “naked-eye” sensor for  $\text{Hg}^{2+}$  in aqueous solution. Other metal ions, such as  $\text{Ag}^+$ ,  $\text{Ba}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ ,

$\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Pb}^{2+}$  and  $\text{Zn}^{2+}$  did not show any significant spectral change except  $\text{Fe}^{3+}$  and  $\text{Cu}^{2+}$  caused a little bit of enhancement under identical conditions (Fig. 3). These results suggested that Rh-C could serve as a “naked-eye” chemosensor selective for  $\text{Hg}^{2+}$  in neutral buffered media.

The method of continuous variations (Job’s plot) obtained from the Rh-C+ $\text{Hg}^{2+}$  system in buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v), When molar fraction of  $\text{Hg}^{2+}$  was 0.5, the absorbance at 562 nm got to maximum (Fig. 2.Inset (b)), indicating that forming a 1:1 complex between Rh-C and  $\text{Hg}^{2+}$ , which was confirmed by the Benesi-Hildebrand method [32].

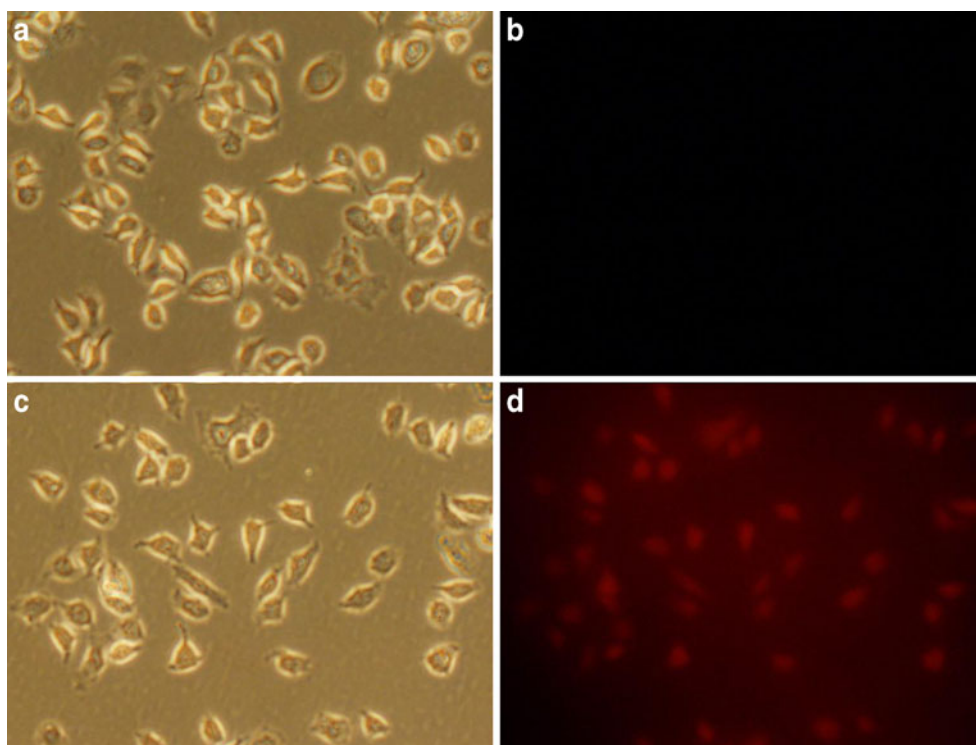


**Scheme 2** Probable complexation mechanism of Rh-C with  $\text{Hg}^{2+}$



**Fig. 8** Competition of Rh-C (10.0  $\mu\text{M}$ ) for  $\text{Hg}^{2+}$  (2equiv.) upon addition of other metal ions (10equiv.) in buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v).  $\Delta F = F - F_0$ , ( $F_0$  and  $F$  were the fluorescence intensity of Rh-C in the absence and presence of  $\text{Hg}^{2+}$ ). Excitation: 535 nm, emission: 586 nm

**Fig. 9** Fluorescence images of  $\text{Hg}^{2+}$  in HL-7702 cells with 5  $\mu\text{M}$  solution of Rh-C in Ethanol-PBS (1 : 99, v/v) buffer for 30 min at 37 °C, Bright-field transmission image (a, c) and fluorescence image (b, d) of HL-7702 cells incubated with 0  $\mu\text{M}$ , 5  $\mu\text{M}$  of  $\text{Hg}^{2+}$  for 30 min, respectively



Linear fitting of the titration profiles using Benesi-Hildebrand plot based on a 1:1 binding mode results in a good linearity (Fig. 4), which strongly support the 1:1 binding stoichiometry of Rh-C and  $\text{Hg}^{2+}$ , and the binding constant was calculate to be  $3.81 \times 10^4 \text{ M}^{-1}$  [33].

#### Fluorescence Spectral Responses of Rh-C

Fluorescence titrations of Rh-C with  $\text{Hg}^{2+}$  in buffered (HEPES 20 mM, pH=7.0) water-ethanol (7/3, v/v) were then performed. As shown in Fig. 5, free Rh-C showed a very weak band in the range 500–700 nm due to the spirocyclic structure. Upon addition gradual increase of  $\text{Hg}^{2+}$  to the solution of Rh-C (10  $\mu\text{M}$ ), a significant enhancement in fluorescence intensity at 586 nm was observed following excitation at 535 nm and gradual increased with  $\text{Hg}^{2+}$  concentrations. Meanwhile the solution showed an orange fluorescence.

Fitting of the Job's plot evaluated from the fluorescent spectra of Rh-C and  $\text{Hg}^{2+}$  at 586 nm gave rise to a 1: 1 stoichiometry for the Rh-C- $\text{Hg}^{2+}$  complex (Fig 5 Inset (b)), which consistent with the results of absorption. The linear response of the fluorescence intensity toward  $\text{Hg}^{2+}$  was obtained in  $\text{Hg}^{2+}$  concentration range of 0.4–5  $\mu\text{M}$  (Fig 5 Inset (a)). And the limit of detect ion (LOD) was obtained of  $7.4 \times 10^{-8} \text{ M}$ , which was calculated based on  $3 \delta/k$  ( $\delta$  is the standard deviation of the measured intensity of the blank solution and  $k$  is the slope of the plot in the inset of Fig. 5). The results

demonstrated that the probe can quantitatively determine  $\text{Hg}^{2+}$  at the environmentally relevant level.

#### The Proposed Reaction Mechanism

In addition, the  $\text{Na}_2\text{S}$ -adding experiments were conducted to examine the reversibility of this reaction as shown in Fig. 6. Firstly, the addition of  $\text{Hg}^{2+}$  to the free Rh-C solution makes the fluorescence increase significantly. Secondly, the addition of  $\text{Na}_2\text{S}$  could immediately restore the initial fluorescent intensity and the color of free Rh-C due to the  $K_d$  value of  $10^{-50} \text{ M}^2$  for  $\text{Hg}^{2+}$  at a standard condition in the form of  $[\text{HgS}_2]^{2-}$ . The stability of Rh-C has been test by add  $\text{Hg}^{2+}$  and  $\text{Na}_2\text{S}$  alternately. After four cycles, the fluorescence intensity of Hg-Rh-C quenched a little compared to the original fluorescent intensity. Thus, the probe Rh-C may be used repeatedly by adding  $\text{Na}_2\text{S}$ . It was also confirmed that the response of Rh-C to  $\text{Hg}^{2+}$  was reversible rather than a cation-catalyzed reaction.

Thus, based on the 1:1 binding mode and the reversible behaviour between Rh-C and  $\text{Hg}^{2+}$ , a possible coordination mode for Rh-C with  $\text{Hg}^{2+}$  was proposed (Scheme 2). Rh-C chelates with  $\text{Hg}^{2+}$  via two O atoms of two carbonyl groups, forming the complexation that requires the opening of the spiroring, which caused the recovery of fluorescence of rhodamine. It is very likely due to the chelation-induced ring opening of rhodamine spirolactam, rather than other possible reactions [25, 34].

## Selectivity and Completion

The selectivity of Rh-C for  $\text{Hg}^{2+}$  was further observed in the fluorescent spectra. As expected, Rh-C exhibited excellent fluorescence selectivity towards  $\text{Hg}^{2+}$  over all other alkali and alkaline earth metal ions, transition and heavy metal ions except for a little bit of fluorescence enhancement for  $\text{Fe}^{3+}$  (Fig. 7). And, for reliable application, we can add  $\text{F}^-$  to mask  $\text{Fe}^{3+}$  caused the fluorescence. This finding indicated that Rh-C could selectively recognize  $\text{Hg}^{2+}$  in ethanol aqueous condition.

Then competitive experiments were conducted in the presence of 2.0 equiv. of  $\text{Hg}^{2+}$  mixed with 10.0 equiv. of various cations in the solution of Rh-C (10  $\mu\text{M}$ ), respectively. No significant variation in fluorescence intensity was found by comparison with that without other metal ions besides  $\text{Hg}^{2+}$  except for  $\text{Ag}^+$  and  $\text{Zn}^{2+}$  caused a little decrease (Fig. 8). All these indicate that the selectivity of Rh-C for  $\text{Hg}^{2+}$  over other competitive cations is high.

## Bioimaging Applications of Rh-C in HL-7702 Cells

Bioimaging applications of Rh-C for monitoring of  $\text{Hg}^{2+}$  ions in living cells were then carried out. HL-7702 cells were incubated with Rh-C (5  $\mu\text{M}$ ) in culture medium for 30 min at 37 °C, and very weak fluorescence of Rh-C inside the living HL-7702 cells was observed (Fig. 9b). After three times washing with PBS buffer, The cells were then supplemented with 5  $\mu\text{M}$   $\text{Hg}(\text{NO}_3)_2$  in the growth medium for another 30 min at 37 °C, a bright fluorescence was observed from the intracellular (Fig. 8d). A bright-field transmission image of cells treated with Rh-C and  $\text{Hg}^{2+}$  confirmed that the cells were viable throughout the imaging experiments (Fig. 9a, c). It is proved that Rh-C is cell-permeable and primarily little toxic to the cell culture. These results demonstrated that Rh-C may be used for detecting  $\text{Hg}^{2+}$  in biological samples.

## Conclusion

In summary, we have developed a novel easily available turn-on fluorescent sensor Rh-C based on a rhodamine conjugate. It selectively responds to  $\text{Hg}^{2+}$  by chromo- and fluorogenic changes and also facilitates “naked-eye” detection of  $\text{Hg}^{2+}$ . The background metal ions showed small or no interference with the detection of  $\text{Hg}^{2+}$  ion. The chemosensor displayed a linear response to  $\text{Hg}^{2+}$  in the range of 0.4–5  $\mu\text{M}$  with a detection limit of  $7.4 \times 10^{-8}$  M. Moreover, fluorescence microscopy experiments establish that Rh-C can be used for detecting  $\text{Hg}^{2+}$  within living cells.

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